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# COX-2 Inhibition, *H. pylori* Infection and the Risk of Gastrointestinal Complications

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**Abstract:** Current data on the gastric safety of cyclooxygenase-2 (COX-2) inhibitors in the presence of *H. pylori* infection are largely derived from animal experiments and indirect clinical evidence. In animal models of *H. pylori* gastritis, COX-2 inhibitors suppressed prostaglandin synthesis and aggravated mucosal damage. In the human stomach, COX-1 appears to be the predominant source of prostaglandins despite the fact that COX-2 is upregulated in *H. pylori* gastritis. There are conflicting data on whether *H. pylori* alters the risk of ulcer in patients receiving COX-2 inhibitors. Among patients with *H. pylori* infection, rofecoxib reduced the risk of complicated gastric but not duodenal ulcers as compared to naproxen. The advantage of rofecoxib over naproxen also disappeared in patients with *H. pylori* infection and prior upper gastrointestinal events. In contrast, pooled data suggested that *H. pylori* increases the risk of ulcer in patients receiving nonselective nonsteroidal anti-inflammatory drugs but not in patients receiving celecoxib. In rodent gastric ulcers, COX-2 was upregulated in the granulation tissue and ulcer margin. Inhibition of COX-2 delayed healing of experimental gastric ulcer. Limited data showed that COX-2 expression was also increased in human gastric ulcer regardless of the *H. pylori* status. The functional significance of COX-2 in human gastric ulcer is unknown.

**Key Words:** *H. pylori*, cyclooxygenase-2, COX-2 inhibitors, prostaglandins, ulcer.

## INTRODUCTION

*Helicobacter pylori* and nonsteroidal anti-inflammatory drugs (NSAIDs) are the two most important causes of gastroduodenal ulcer disease worldwide. Since many NSAID users are infected with *H. pylori*, it is important to determine whether *H. pylori* would influence the risk of developing ulcers in these patients. It is generally thought that *H. pylori* and NSAIDs are independent risk factors for ulcer disease because they damage the gastric mucosa via different mechanisms. *H. pylori* induces proinflammatory cytokines, leading to mucosal inflammation and epithelial injury. In contrast, NSAIDs damage the gastric mucosa by inhibiting gastric prostaglandin synthesis. However, this view may be simplistic because *H. pylori* and NSAIDs share certain pathways in the pathogenesis of mucosal injury [1, 2]. The controversy about the role of *H. pylori* in NSAID-associated gastroduodenal damage hinges on whether the effects of *H. pylori* and NSAIDs on gastric mucosal damage is synergistic, additive, or antagonistic, and whether there is sufficient clinical evidence to draw any conclusion. Current data suggest that *H. pylori* infection probably has a diverse effect on the gastric mucosa in different subgroups of NSAID users, which partly accounts for the conflicting results on the interaction between *H. pylori* and NSAIDs in mucosal damage [1, 2].

Development of NSAIDs that selectively inhibit cyclooxygenase-2 (COX-2) offers the prospect of relieving pain and inflammation without inflicting gastric injury. In healthy volunteers, selective inhibition of COX-2 does not

suppress gastric prostaglandins [3]. There is good clinical evidence that COX-2 specific inhibitors cause fewer clinical upper gastrointestinal events compared with nonselective NSAIDs [4, 5]. However, the gastrointestinal safety of COX-2 specific inhibitors in the presence of mucosal inflammation remains unclear. COX-2 is induced in gastrointestinal inflammatory conditions, such as inflammatory bowel disease and *H. pylori* gastritis. Inhibition of COX-2 has been shown to suppress colonic prostaglandin synthesis in ulcerative colitis and Crohn's disease [6, 7]. In the rat colitis model, COX-2 specific inhibitor exacerbates colonic inflammation [8]. In the stomach, *H. pylori* induces mucosal inflammation and has been shown to upregulate the expression of COX-2 [1, 6, 7, 9, 10]. This raises the possibility that COX-2 may be the predominant source of prostaglandins in *H. pylori* gastritis, leading to an increased susceptibility to mucosal injury by COX-2 specific inhibitors. To date there are conflicting data showing that COX-2 specific inhibitors increase or have no effect on the risk of mucosal injury in the presence of *H. pylori* gastritis. How COX-2 specific inhibitors differ from nonselective NSAIDs in terms of their effects on *H. pylori*-infected gastric mucosa will be an interesting area of research.

## Expression and Cellular Localization of COX-2 in *H. pylori* Infection

Many studies have reported an increased expression of COX-2 in the presence of *H. pylori* infection. *H. pylori* has been shown to upregulate the expression of COX-2 messenger RNA (mRNA) and stimulates prostaglandin synthesis in gastric cancer cell lines [11]. However, there are conflicting data on the cellular localization of COX-2 expression in *H. pylori* gastritis. Fu *et al.* reported that *H. pylori* induces COX-2 expression in the mononuclear

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inflammatory cells and myofibroblasts in the lamina propria [10]. However, other studies found that COX-2 was expressed mainly in the gastric epithelium [1, 6, 7]. Sawaoka *et al.* reported that COX-2 was expressed both in the gastric epithelium and subepithelial inflammatory cells in *H. pylori* gastritis [9]. These inconsistent immunohistochemical findings probably are related to different laboratory conditions and cross-reactivity of COX-2 antibodies with other mucosal antigens. Using *in situ* hybridization, it has been shown that *H. pylori* up-regulates the expression of COX-2 mRNA mainly in the gastric epithelial cells [1].

#### Role of COX-1 and COX-2 in *H. pylori*-Induced Prostaglandin Synthesis

In the normal human stomach, COX-2 is absent or minimally expressed whereas COX-1 is the source of prostaglandins that maintains the integrity of the mucosal barrier. This notion is consistent with the observation that in the absence of *H. pylori* infection, inhibition of COX-2 did not suppress gastric prostaglandin synthesis and inflicted minimal mucosal injury [3]. However, in cultured human gastric fibroblasts [12, 13], *H. pylori* induced the expression of COX-2 mRNA and increased prostaglandin synthesis. Indomethacin and a COX-2 inhibitor (NS-398) suppressed *H. pylori*-induced prostaglandin synthesis to the same extent. These findings suggested that COX-2 may substantially contribute to prostaglandin synthesis in *H. pylori* gastritis, and that selective inhibition of COX-2 may lose its gastric sparing effect in the presence of *H. pylori* infection.

However, there were conflicting data on the relative contributions of COX-1 and COX-2 in prostaglandin synthesis associated with *H. pylori* gastritis. Jackson *et al.* [14] reported that COX-1 and COX-2 were constitutively expressed in parietal cells of uninfected human stomach. Immunostaining for both COX-1 and COX-2 was increased in *H. pylori* gastritis. Interestingly, the increased *ex vivo* prostaglandin synthesis was significantly suppressed by a COX-1 inhibitor rather than a COX-2 inhibitor. Scheiman *et al.* studied the effect of rofecoxib on gastric prostaglandin synthesis in subjects with or without *H. pylori* infection [15]. Twenty *H. pylori*-infected and 6 uninfected healthy volunteers were treated with rofecoxib for 2 weeks. Although prostaglandin levels were increased in *H. pylori* gastritis, rofecoxib did not suppress prostaglandin synthesis in infected subjects. These results suggested that despite an upregulation of COX-2, *H. pylori* gastritis, COX-1 remains the predominant source of prostaglandins.

#### EFFECTS OF NSAIDS AND COX-2 SPECIFIC INHIBITORS ON *H. pylori*-INFECTED GASTRIC MUCOSA

Gastric prostaglandins play a crucial role in mucosal defense by regulating mucosal blood flow, mucus and bicarbonate secretion, epithelial proliferation, epithelial restitution, and mucosal immunocyte function [16]. The fact that *H. pylori* infection stimulates gastric prostaglandin production has led to the speculation that *H. pylori* may alleviate mucosal injury induced by NSAIDs. However, administration of NSAIDs to *H. pylori*-infected subjects has been shown to profoundly suppress prostaglandin production to levels that were similar to those of uninfected subjects [17, 18]. These findings indicate that the modest increase in prostaglandin levels induced by *H. pylori* is unlikely to have any important protective effect against NSAID injury. It has been postulated that the mucosal toxicity of *H. pylori*, which is largely mediated by inflammatory cytokines including interleukin (IL)-1, IL-6, IL-8 and tumor necrosis factor, is counterbalanced by protective responses, such as increased release of mucosal prostaglandins and hepatocyte growth factor [12]. Factors disturbing this balance may enhance the gastric damaging effects of NSAIDs and mucosal toxicity of *H. pylori* [12]. To date there are only a few experimental and human studies investigating the effects of selective COX-2 inhibition on *H. pylori*-infected gastric mucosa.

#### Animal Studies (Table 1)

Several studies have investigated the effect of nonselective NSAIDs and a COX-2 specific inhibitor on *H. pylori* gastritis using the Mongolian gerbil model [13, 19, 20]. In one study [13], COX-1 was detected in both normal and *H. pylori*-infected mucosa whereas COX-2 was expressed only in the infected mucosa. *H. pylori* infection increased prostaglandin synthesis. Indomethacin markedly inhibited prostaglandin synthesis in both normal and infected mucosa. NS-398 also suppressed prostaglandin synthesis in infected mucosa but did not have any effect on uninfected mucosa. Hemorrhagic erosions and neutrophil infiltration were found in *H. pylori* gastritis. These mucosal lesions were aggravated by indomethacin and NS-398. Both drugs potentiated the release of neutrophil chemokine and interferon- $\gamma$  induced by *H. pylori*. In another study [19], indomethacin and NS-398 significantly suppressed gastric prostaglandin synthesis and there was a non-significant trend toward less severe suppression with NS-398 in *H. pylori*-infected gerbils. Indomethacin and NS-398 caused similar degree of gastric mucosal damage in infected animals despite

Table 1. Effects of NSAIDs and COX-2 Inhibitor on Animal Models of *H. pylori* Gastritis

Animal model	COX-1 & -2 expression	Baseline PGE2	PGE2 after NSAIDs	PGE2 after COX-2 inhibitor
Mongolian Gerbil [13]	COX-2 upregulated	Increased	Suppressed	Moderately suppressed*
Mongolian Gerbil [19]	---	Increased	Suppressed	Moderately suppressed†
Mouse [21]	COX-1 & -2 upregulated	Non-significant increase	Suppressed	Non-significant decrease

\*Mucosal damage was aggravated by both NSAIDs and NS-398 in *H. pylori*-infected mucosa.

†NSAIDs and NS-398 induced similar degree of mucosal damage in *H. pylori*-infected animals.

different degrees of prostaglandin suppression. In contrast, there was an inverse relationship between gastric prostaglandin level and mucosal damage in uninfected animals. These findings suggested that while COX-2 specific inhibitors caused minimal injury to uninfected gastric mucosa, these drugs did not reduce mucosal damage in *H. pylori* gastritis.

Unlike the previous two studies, Kim *et al.* [21] found that both COX-1 and COX-2 were upregulated in mouse stomachs infected with *H. pylori*. *H. pylori* infection increased apoptotic index, cell proliferation index, neutrophil activity and the degree of chronic inflammation. There was a non-significant increase in gastric prostaglandin levels. All these changes were reversed after the administration of indomethacin whereas NS-398 did not induce a significant reduction. The result suggested that both COX-1 and COX-2 are induced by *H. pylori* infection. Induction of COX-1 also contributes to the increase in prostaglandin synthesis, mucosal cell turnover and inflammatory activity in *H. pylori* gastritis.

Recently, Futagami *et al.* [20] investigated how inhibition of COX-2 would influence the severity of NSAID-induced gastric damage in *H. pylori*-infected Mongolian gerbils. *H. pylori* infection induced COX-2 expression. Prolonged treatment with indomethacin caused more severe gastric damage in *H. pylori*-infected animals than in uninfected animals. Interestingly, pretreatment with NS-398 aggravated the mucosal damage induced by short-term treatment with indomethacin in the presence of *H. pylori*. The authors postulated that induction of COX-2 by *H. pylori* might protect the gastric mucosa against NSAID injury. Disturbance of this equilibrium state by inhibiting COX-2 may enhance the gastric toxicity of NSAIDs in *H. pylori*-infected animals (Fig. 1).

#### Human Studies (Table 2)

Current evidence on the gastric safety of COX-2 inhibitors in *H. pylori*-infected patients is mostly derived

from *post hoc* analysis. Whether *H. pylori* infection increases the risk of ulcer disease in patients receiving COX-2 specific inhibitors has generated conflicting results in the literature.

The influence of *H. pylori* infection on the risk of gastroduodenal ulceration was first reported in a subgroup analysis of a double-blind, 12-week endoscopic study of celecoxib versus naproxen [22]. Among patients who received celecoxib, the incidence of ulcer was 12.9% in patients with *H. pylori* infection compared with 2.9% in uninfected patients ( $P=0.023$ ). Other risk factors included concurrent use of low-dose aspirin ( $P=0.001$ ) and a history of ulcer ( $P=0.010$ ). In contrast, *H. pylori* did not influence the risk of ulcer among patients who received naproxen. However, the same group of investigators reported contradictory results in a pooled analysis of four double-blind 12-week endoscopic studies of celecoxib that collectively enrolled 4000 arthritis patients [23]. Among patients who used nonselective NSAIDs, the incidence of ulcer was 28.4% in patients with *H. pylori* infection compared with 20% in uninfected patients (odds ratio 1.6 [1.1, 2.3]). Among patients who used celecoxib, the incidence of ulcer was 8.0% in patients with *H. pylori* infection compared with 5.1% in uninfected patients (odds ratio 1.6 [0.9, 2.8]). These results suggested that *H. pylori* is a risk factor for gastroduodenal ulceration in patients taking nonselective NSAIDs but not celecoxib.

In a multivariate analysis of risk factors for upper gastrointestinal clinical events [24] based on the data collected in the Vioxx Gastrointestinal Outcomes Research Study [5], major risk factors for the development of upper gastrointestinal clinical events included old age ( $\geq 75$ ) and prior complicated or uncomplicated gastrointestinal events. Patients with prior gastrointestinal events who received naproxen had a high rate of clinical events regardless of *H. pylori* status. Although *H. pylori* was not considered as a risk factor in this multivariate analysis, two interesting findings were reported. First, patients in the rofecoxib group had

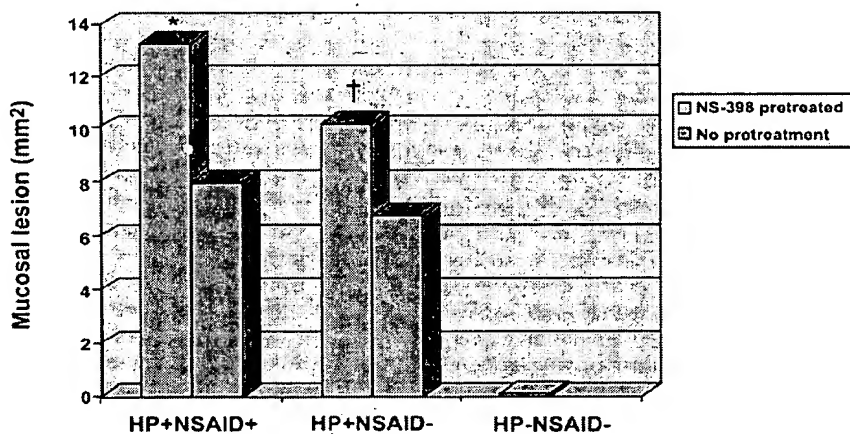


Fig. (1). Effects of Indomethacin on gastric damage in *H. pylori*-infected Mongolian gerbils with or without pretreatment with NS-398. HP and NSAID denote *H. pylori* and indomethacin, respectively. \* $P<0.05$  versus HP+NSAID+ group without pretreatment with NS-398. † $P<0.05$  versus HP+NSAID- group without pretreatment with NS-398. Data derived from [20].

Table 2. Clinical Effects of *H. pylori* Infection on Gastroduodenal Damage of COX-2 Specific Inhibitors

Design	Number of patients	Outcomes				
Subgroup analysis of a 12-week RCT of celecoxib 400 mg versus naproxen 1 g [22]	536	Endoscopic ulcer:				
			Celecoxib	Naproxen		
		HP positive	12.9%*	29%		
		HP negative	2.9%	30%		
		(P=0.023)*				
Pooled analysis of four 12-week RCTs of celecoxib 100-800 mg versus nonselective NSAIDs [23]	4000	Endoscopic ulcer (non-aspirin users):				
			Celecoxib	Nonselective NSAIDs		
		HP positive	8.0%	28.4%		
		HP negative	5.1%	20.0%		
		OR	1.6 (0.9 - 2.8)	1.6 (1.1 – 2.3)*		
Multivariate analysis of a RCT of rofecoxib 50 mg versus naproxen 1 g [24]	8076	Clinical upper GI events (per 100 patient-years):				
			Rofecoxib	Naproxen		
		Prior event, HP positive	12.18	14.00	RR 0.89 (0.38-2.07)†	
		Prior event, HP negative	3.35	17.14	RR 0.20 (0.07-0.61)	
		Complicated DU (per 100 patient-years):				
			Rofecoxib	Naproxen		
		HP positive	1.85	1.54	RR 1.20 (0.64-2.24)‡	
		HP negative	0.34	1.41	RR 0.24 (0.09-0.64)	

\**H. pylori* was a risk factor in patients taking nonselective NSAIDs but not in patients taking celecoxib

†The upper GI sparing effect of rofecoxib was offset by the presence of *H. pylori* infection in patients with prior upper GI events.

‡The superiority of rofecoxib over naproxen in reducing the risk of complicated duodenal ulcers disappeared in the presence of *H. pylori* infection.

fewer gastric ulcers than patients in the naproxen group regardless of the *H. pylori* status. In contrast, rofecoxib did not reduce the risk of duodenal ulcers compared with naproxen among patients found positive for *H. pylori*. Second, among those with prior gastrointestinal events, the rate of events in the rofecoxib group was 3.5-fold higher in *H. pylori*-positive patients than in *H. pylori*-negative patients. The results suggested that the upper GI sparing effect of rofecoxib was the offset by the presence of *H. pylori* infection in patients with prior upper GI events, and the superiority of rofecoxib over naproxen in reducing the risk of complicated duodenal ulcers disappeared in the presence of *H. pylori* infection.

#### EFFECTS OF *H. pylori* ON ULCER HEALING ASSOCIATED WITH NSAIDS AND COX-2 SPECIFIC INHIBITORS

Whether *H. pylori* infection affects ulcer healing in patients receiving nonselective NSAIDs has yielded conflicting data. In the rat model, one study found that *H. pylori* attenuated the delay in ulcer healing induced by aspirin. This observation was attributed to suppression of acid secretion and stimulation of prostaglandin production by *H. pylori* [25]. However, there are conflicting findings about the effects of *H. pylori* on aspirin-induced gastric injury. The same group of investigators showed that *H. pylori* induced persistent mucosal bleeding in the rat stomach by impairing gastric adaptation to aspirin [18]. Eradication of *H. pylori*

restores gastric adaptation to resist aspirin-induced injury. Hawkey *et al.* studied the effect of *H. pylori* eradication on gastroduodenal damage in chronic NSAID users with dyspepsia or ulcer [26]. In a subgroup of 41 patients with gastric ulcers, they found that eradication of *H. pylori* delayed ulcer healing (ulcer healing at 8 weeks: 72% in the eradicated group compared with 100% in the control group). In another randomized trial of *H. pylori*-positive patients with NSAID-associated ulcer bleeding, 195 patients (112 gastric ulcers and 83 duodenal ulcers) were randomly assigned to receive omeprazole or omeprazole plus eradication therapy for ulcer healing. Eradication of *H. pylori* did not have any significant adverse effect on the healing of gastric (*H. pylori*-positive versus *H. pylori*-eradicated: 94% versus 88%;  $p=0.29$ ) or duodenal (*H. pylori*-positive versus *H. pylori*-eradicated: 100% versus 98%;  $p=1.0$ ) ulcers [27] (Fig. 2). To date there is no definite evidence to show that eradication of *H. pylori* has any clinically important adverse effect on healing of NSAID-associated ulcers.

On the other hand, there are data suggesting that among patients receiving NSAIDs, gastric ulcers heal faster with *H. pylori* infection by acid suppression [28, 29]. *H. pylori* infection has been shown to augment the acid-suppressing effect of omeprazole [30, 31]. In a pooled analysis of three randomized trials of omeprazole for the prevention of mucosal injury in NSAID users [28], *H. pylori* appeared to enhance gastric ulcer healing by acid suppression but retard

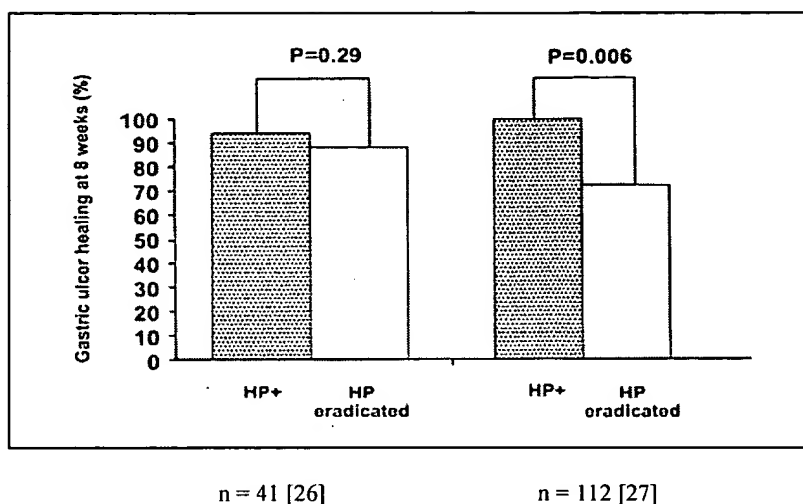


Fig. (2). Effects of *H. pylori* eradication on healing of gastric ulcers in patients receiving nonselective NSAIDs.

healing by misoprostol. However, the difference only reached statistical significance in patients receiving ranitidine (84% vs 51% at eight weeks) but not in patients receiving omeprazole. In another pooled analysis of two randomized trials of lansoprazole versus ranitidine in preventing NSAID-induced mucosal injury [29], gastric ulcer healing rates were significantly faster in *H. pylori*-positive patients than in uninfected patients though the difference was of doubtful clinical relevance (70% vs 61% at 8 weeks;  $P < 0.05$ ).

Although COX-2 specific inhibitors inflict minimal gastric injury to the normal stomach, there is evidence from animal experiments that COX-2 may play a physiological role in restoring mucosal integrity. In the rat stomach, COX-2 was upregulated in acetic acid-induced gastric ulcers [32, 33]. COX-2 activity was detected in endothelial cells, macrophages and fibroblasts at the ulcer base [32, 33]. Selective inhibition of COX-2 has been shown to retard gastric ulcer healing [33-36]. Administration of NS-398 significantly retarded healing of acetic acid-induced ulcers in rats and thermal-cauterized ulcers in mice [33-35]. One study showed that inhibition of COX-2-derived prostaglandins in ulcerated mucosa delayed ulcer healing [33]. Other investigators found that angiogenesis and maturation of granulation tissue in gastric ulcer was impaired by inhibition of COX-2 [36]. Jones *et al.* [37] demonstrated that both nonselective NSAIDs and COX-2 inhibitors acted on endothelial cells to inhibit angiogenesis. In contrast, COX-1 was absent [36] or not induced in ulcerated mucosa [33, 34].

Unlike rodent ulcers, To *et al.* [38] found that both COX-1 and COX-2 were upregulated in human gastric ulcers. At the ulcer margin, increased COX-1 expression was detected in lamina propria cells whereas COX-2 was strongly expressed in the hyperplastic foveolar epithelium. At the ulcer base, COX-1 and COX-2 were strongly expressed in myofibroblasts, macrophages and endothelial cells in the

granulation tissue. The findings were similar between *H. pylori* ulcers or NSAIDs ulcers. This raises the possibility that both isoforms of COX may contribute to ulcer healing in the human stomach regardless of the *H. pylori* status. However, other investigators found that although intense COX-2 immunoreactivity was detected in human gastric ulcers, there was no significant change in COX-1 expression in ulcerated mucosa [14, 39]. To date there is no clinical data as to whether COX-2 inhibitors would retard gastric ulcer healing.

## CONCLUSION

Current data on the interaction between *H. pylori* infection and selective COX-2 inhibition with respect to gastric damage are mostly derived from animal experiments or indirect clinical evidence based on *post hoc* analysis. Several interesting findings deserve further studying. In animal models of *H. pylori* gastritis, COX-2 inhibitors suppressed prostaglandin synthesis and aggravated mucosal damage. COX-1 appears to be the predominant source of prostaglandins in the human stomach albeit an upregulation of COX-2 in the presence of *H. pylori* infection [14, 15]. This may partly explain why *H. pylori* did not increase the risk of developing gastric ulcers among patients receiving rofecoxib [24]. However, the same study also indicated that rofecoxib did not reduce the risk of complicated duodenal ulcers in the presence of *H. pylori* infection. In addition, there was no advantage of rofecoxib over a nonselective NSAID for those with prior events and *H. pylori* infection in terms of the risk of clinical upper gastrointestinal events. Whether eradication of *H. pylori* will reduce the ulcer risk in these subgroups has not been investigated. In contrast, pooled analysis of data from randomized trials of celecoxib showed that *H. pylori* was a risk for ulcer disease in patients receiving nonselective NSAIDs but not in patients receiving celecoxib [23]. It is uncertain whether these contradictory

findings reflect differences in pharmacological properties, variations in study design or heterogeneity of *H. pylori*-infected patients. In animal models of gastric ulcer, COX-2 was upregulated in the granulation tissue and ulcer margin. Inhibition of COX-2 delayed healing of gastric ulcer in rodents. Limited data showed that COX-2 expression was increased in human gastric ulcer regardless of the *H. pylori* status. Whether inhibition of COX-2 will impair ulcer healing in the human stomach remains unknown. Future studies with pre-specified endpoints are needed to define the gastrointestinal risk of COX-2 inhibitors in different subgroups of *H. pylori*-infected patients.

## REFERENCES

References 40-42 are related articles recently published in *Current Pharmaceutical Design*.

- [1] Chan FK, To KF, Ng YP, Lee TL, Cheng AS, Leung WK et al. Expression and cellular localization of cyclooxygenase -1 and -2 in *Helicobacter pylori* gastritis. *Aliment Pharmacology Ther* 2001; 15: 187-194.
- [2] Chan FK. *Helicobacter pylori* and nonsteroidal anti-inflammatory drugs. *Gastroenterology Clin North Am* 2001; 30: 937-952.
- [3] Wight NJ, Gottesdiener K, Garlick NM, Atherton CT, Novak S, Gertz BJ, et al. Rofecoxib, a COX-2 inhibitors, does not inhibit human gastric mucosal prostaglandin production. *Gastroenterology* 2001; 120: 867-73.
- [4] Silverstein FE, Faich G, Goldstein JL, Simon LS, Pincus T, Whelton A, et al. Gastrointestinal toxicity with celecoxib vs nonsteroidal anti-inflammatory drugs for osteoarthritis and rheumatoid arthritis: the CLASS study: A randomized controlled trial. Celecoxib Long-term Arthritis Safety Study. *JAMA* 2000; 284: 1247-1255.
- [5] Bombardier C, Laine L, Reicin A, Shapiro D, Burgos-Vargas R, Davis B, et al. Comparison of upper gastrointestinal toxicity of rofecoxib and naproxen in patients with rheumatoid arthritis. *N Engl J Med* 2000; 343: 1520-1528.
- [6] McCarthy CJ, Crofford LJ, Greenon J, Scheiman JM. Cyclooxygenase-2 expression in gastric antral mucosa before and after eradication of *Helicobacter pylori* infection. *Am J Gastroenterology* 1999; 94: 1218-23.
- [7] McCartney SA, Mitchell JA, Fairclough PD, Farthing MJ, Warner TD. Selective COX-2 specific inhibitors and human inflammatory bowel disease. *Aliment Pharmacol Ther* 1999; 13: 115-117.
- [8] Reuter BK, Asfaha S, Buret A, Sharkey KA, Wallace JL. Exacerbation of inflammation-associated colonic injury in rat through inhibition of cyclooxygenase-2. *J Clin Invest* 1996; 98: 2076-2085.
- [9] Sawaoka H, Kawano S, Tsuji S, Tsuji M, Sun W, Gunawan ES, et al. *Helicobacter pylori* infection induces cyclooxygenase-2 expression in human gastric mucosa. *Prostaglandins Leukot Essent Fatty Acids* 1998; 59: 313-316.
- [10] Fu S, Ramanujam KS, Wong A, Fantry GT, Drachenberg CB, James SP, et al. Increased expression and cellular localization of inducible nitric oxide synthase and cyclooxygenase 2 in *Helicobacter pylori* gastritis. *Gastroenterology* 1999; 116: 1319-29.
- [11] Romano M, Ricci V, Memoli A, Tuccillo C, Di Popolo A, Sommi P, et al. *Helicobacter pylori* up-regulates cyclooxygenase-2 mRNA expression and prostaglandin E<sub>2</sub> synthesis in MKN 28 gastric mucosal cells in vitro. *J Biology Chem* 1998; 273: 28560-28563.
- [12] Takahashi M, Katayama Y, Takada H, Kuwayama H, Terano A. The effect of NSAIDs and a COX-2 specific inhibitor on *Helicobacter pylori*-induced PGE<sub>2</sub> and HGF in human gastric fibroblasts. *Aliment Pharma Ther* 2000; 14; (Suppl 1), 44-49.
- [13] Takahashi S, Fujita T, Yamamoto A. Role of cyclooxygenase-2 in *Helicobacter pylori*-induced gastritis in Mongolian gerbils. *Am J Physiology Gastrointest Liver Physiology* 2000; 279: G791-G798.
- [14] Jackson LM, Wu KC, Mahida YR, Jenkins D, Hawkey CJ. Cyclooxygenase (COX) 1 and 2 in normal, inflamed, and ulcerated human gastric mucosa. *Gut* 2000; 47: 762-770.
- [15] Scheiman JM, Greenon JK, Lee J, Cryer B. Impact of COX-2 specific inhibition on human *Helicobacter pylori* (HP) Gastritis: Implications for ulcerogenesis and carcinogenesis. *Gastroenterology* 2002; 122: A87.
- [16] Wallace JL, Tigley AW. New insights into prostaglandins and mucosal defence. *Aliment Pharmacology Ther* 1995; 9: 227-235.
- [17] Laine L, Cominelli F, Sloane R, Casini-Raggi V, Marin-Sorensen M, Weinstein WM. Interaction of NSAIDs and *Helicobacter pylori* on gastroduodenal injury and prostaglandin production: a controlled double-blind trial. *Aliment Pharmacology Ther* 1995; 9: 127-35.
- [18] Konturek JW, Dembinski A, Konturek SJ, Stachura J, Domschke W. Infection of *Helicobacter pylori* in gastric adaptation to continued administration of aspirin in humans. *Gastroenterology* 1998; 114: 245-255.
- [19] Bhang CS, Lee HS, Kim SS, Song HJ, Sung YJ, Kim JI, et al. Effects of selective cyclooxygenase-2 inhibitor and non-selective NSAIDs on *Helicobacter pylori*-induced gastritis in Mongolian gerbils. *Helicobacter* 2002; 7: 14-21.
- [20] Futagami S, Hiratsuka T, Wada K, Tatsuguchi A, Tsukui T, Miyake K, et al. Inhibition of *Helicobacter pylori*-induced cyclooxygenase-2 aggravates NSAID-caused gastric damage in Mongolian gerbils. *Aliment Pharmacology Ther* 2002; 16: 847-855.
- [21] Kim TI, Lee YC, Lee KH, Han JH, Chon CY, Moon YM, et al. Effects of nonsteroidal anti-inflammatory drugs on *Helicobacter pylori*-infected gastric mucosae of mice: apoptosis, cell proliferation, and inflammatory activity. *Infect Immun* 2001; 69: 5056-63.
- [22] Goldstein JL, Correa P, Zhao WW, Burr AM, Hubbard RC, Verburg KM, et al. Reduced incidence of gastroduodenal ulcers with celecoxib, a novel cyclooxygenase-2 inhibitor, compared to naproxen in patients with arthritis. *Am J Gastroenterology* 2001; 96: 1019-1027.
- [23] Goldstein JL, Agrawal NM, Silverstein FE, Verburg KM, Burr AM, Hubbard RC, et al. Influence of *H. pylori* infection and/or low-dose aspirin on gastroduodenal ulceration in patients treated with placebo, celecoxib, or NSAIDs. *Gastroenterology* 1999; 116: A174.
- [24] Laine L, Bombardier C, Hawkey CJ, Davis B, Shapiro D, Brett C, et al. Stratifying the risk of NSAID-related upper gastrointestinal clinical events: results of a double-blind outcomes study in patients with rheumatoid arthritis. *Gastroenterology* 2002; 213: 1006-1012.
- [25] Konturek PC, Brzozowski T, Kwiecien S, Drozdowicz D, Harsch IA, Meixner H, et al. Effect of *Helicobacter pylori* on delay in ulcer healing induced by aspirin in rats. *Eur J Pharmacology* 2002; 451: 191-202.
- [26] Hawkey CJ, Tulassay Z, Szczepanski L, van Rensburg CJ, Filipowicz-Sosnowska A, Lanas A, et al. Randomised controlled trial of *Helicobacter pylori* eradication in patients on non-steroidal anti-inflammatory drugs: HELP NSAIDs study. *Lancet* 1998; 352: 1016-1021.
- [27] Chan FK, Sung JJ, Suen R, Lee YT, Wu JC, Leung WK, et al. Does eradication of *H. pylori* impair healing of nonsteroidal anti-inflammatory drug associated bleeding peptic ulcers? A prospective randomized study. *Aliment Pharmacology Ther* 1998; 12: 1201-1205.
- [28] Hawkey CJ, Wilson I, Naesdal J, Langstrom G, Swannell AJ, Yeomans ND. Influence of sex and *Helicobacter pylori* on development and healing of gastroduodenal lesions in non-steroidal anti-inflammatory drug users. *Gut* 2002; 51: 344-50.
- [29] Campbell DR, Haber MM, Sheldon E, Collis C, Lukasik N, Huang B, et al. Effect of *H. pylori* status on gastric ulcer healing in patients continuing nonsteroidal anti-inflammatory therapy and receiving treatment with lansoprazole or ranitidine. *Am. J. Gastroenterol.* 2002; 97: 2208-14.
- [30] Labenz J, Tillenburger B, Peitz U, Idstrom JP, Verdu EF, Stolte M, et al. *Helicobacter pylori* augments the pH-increasing effect of omeprazole in patients with duodenal ulcer. *Gastroenterology* 1996; 110: 725-732.
- [31] Gillen D, Wirz AA, Neithercut WD, Ardill JES, McCall KEL. *Helicobacter pylori* infection potentiates the inhibition of gastric acid secretion by omeprazole. *Gut* 1999; 44: 468-475.
- [32] Takahashi S, Shigeta J, Inoue H, Tanabe T, Okabe S. Localization of cyclooxygenase-2 and regulation of its mRNA expression in gastric ulcers in rats. *Am J Physiology* 1998; 275: G1137-G1145.

- [33] Shigeta J, Takahashi S, Okabe, S. Role of cyclooxygenase-2 in the healing of gastric ulcers in rats. *J Pharmacology Exp Ther* 1998; 286: 1383-1390.
- [34] Mizuno H, Sakamoto C, Matsuda K, Wada K, Uchida T, Noguchi H, et al. Induction of cyclooxygenase 2 in gastric mucosal lesions and its inhibition by the specific antagonist delays healing in mice. *Gastroenterology* 1997; 112: 387-397.
- [35] Ukawa H, Yamakuni H, Kato S, Takeuchi K. Effects of cyclooxygenase-2 selective and nitric oxide-releasing nonsteroidal antiinflammatory drugs on mucosal ulcerogenic and healing responses of the stomach. *Dig Dis Sci* 1998; 43: 2003-2011.
- [36] Schmassmann A, Peskar BM, Stettler C, Netzer P, Stroff T, Flogerzi B, et al. Effects of inhibition of prostaglandin endoperoxide synthase-2 in chronic gastro-intestinal ulcer models in rats. *Br J Pharmacology* 1998; 123: 795-804.
- [37] Jones MK, Wang H, Peskar BM, Levin E, Itani RM, Sarfeh IJ, et al. Inhibition of angiogenesis by nonsteroidal anti-inflammatory drugs: insight into mechanisms and implications for cancer growth and ulcer healing. *Nat Med* 1999; 5: 1418-1423.
- [38] To KF, Chan FK, Cheng AS, Lee TL, Ng YP, Sung JJ. Up-regulation of cyclooxygenase-1 and -2 in human gastric ulcer. *Aliment Pharmacology Ther* 2001; 15: 25-34.
- [39] Tatsuguchi A, Sakamoto C, Wada K, Akamatsu T, Tsukui T, Miyake K, et al. Localisation of cyclooxygenase 1 and cyclooxygenase 2 in *Helicobacter pylori* related gastritis and gastric ulcer tissues in humans. *Gut* 2000; 46: 782-789.
- [40] Hawkey CJ, Skelly MM. Gastrointestinal safety of selective COX-2 inhibitors. *Curr Pharm Design* 2002; 8(12): 1077-89.
- [41] Legrain P, Strosberg D. Protein interaction domain mapping for the selection of validated targets and lead compounds in the anti-infectious area. *Curr Pharm Design* 2002; 8(13): 1189-98.
- [42] Sobal G, Sinzinger H. Prostaglandins and lipid modification. *Curr Pharm Design* 2001; 7(6): 461-74.



## COX-2 Inhibition, *H. pylori* Infection and the Risk of Gastrointestinal Complications

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**Abstract:** Current data on the gastric safety of cyclooxygenase-2 (COX-2) inhibitors in the presence of *H. pylori* infection are largely derived from animal experiments and indirect clinical evidence. In animal models of *H. pylori* gastritis, COX-2 inhibitors suppressed prostaglandin synthesis and aggravated mucosal damage. In the human stomach, COX-1 appears to be the predominant source of prostaglandins despite the fact that COX-2 is upregulated in *H. pylori* gastritis. There are conflicting data on whether *H. pylori* alters the risk of ulcer in patients receiving COX-2 inhibitors. Among patients with *H. pylori* infection, rofecoxib reduced the risk of complicated gastric but not duodenal ulcers as compared to naproxen. The advantage of rofecoxib over naproxen also disappeared in patients with *H. pylori* infection and prior upper gastrointestinal events. In contrast, pooled data suggested that *H. pylori* increases the risk of ulcer in patients receiving nonselective nonsteroidal anti-inflammatory drugs but not in patients receiving celecoxib. In rodent gastric ulcers, COX-2 was upregulated in the granulation tissue and ulcer margin. Inhibition of COX-2 delayed healing of experimental gastric ulcer. Limited data showed that COX-2 expression was also increased in human gastric ulcer regardless of the *H. pylori* status. The functional significance of COX-2 in human gastric ulcer is unknown.

**Key Words:** *H. pylori*, cyclooxygenase-2, COX-2 inhibitors, prostaglandins, ulcer.

### INTRODUCTION

*Helicobacter pylori* and nonsteroidal anti-inflammatory drugs (NSAIDs) are the two most important causes of gastroduodenal ulcer disease worldwide. Since many NSAID users are infected with *H. pylori*, it is important to determine whether *H. pylori* would influence the risk of developing ulcers in these patients. It is generally thought that *H. pylori* and NSAIDs are independent risk factors for ulcer disease because they damage the gastric mucosa via different mechanisms. *H. pylori* induces proinflammatory cytokines, leading to mucosal inflammation and epithelial injury. In contrast, NSAIDs damage the gastric mucosa by inhibiting gastric prostaglandin synthesis. However, this view may be simplistic because *H. pylori* and NSAIDs share certain pathways in the pathogenesis of mucosal injury [1, 2]. The controversy about the role of *H. pylori* in NSAID-associated gastroduodenal damage hinges on whether the effects of *H. pylori* and NSAIDs on gastric mucosal damage is synergistic, additive, or antagonistic, and whether there is sufficient clinical evidence to draw any conclusion. Current data suggest that *H. pylori* infection probably has a diverse effect on the gastric mucosa in different subgroups of NSAID users, which partly accounts for the conflicting results on the interaction between *H. pylori* and NSAIDs in mucosal damage [1, 2].

Development of NSAIDs that selectively inhibit cyclooxygenase-2 (COX-2) offers the prospect of relieving pain and inflammation without inflicting gastric injury. In healthy volunteers, selective inhibition of COX-2 does not

suppress gastric prostaglandins [3]. There is good clinical evidence that COX-2 specific inhibitors cause fewer clinical upper gastrointestinal events compared with nonselective NSAIDs [4, 5]. However, the gastrointestinal safety of COX-2 specific inhibitors in the presence of mucosal inflammation remains unclear. COX-2 is induced in gastrointestinal inflammatory conditions, such as inflammatory bowel disease and *H. pylori* gastritis. Inhibition of COX-2 has been shown to suppress colonic prostaglandin synthesis in ulcerative colitis and Crohn's disease [6, 7]. In the rat colitis model, COX-2 specific inhibitor exacerbates colonic inflammation [8]. In the stomach, *H. pylori* induces mucosal inflammation and has been shown to upregulate the expression of COX-2 [1, 6, 7, 9, 10]. This raises the possibility that COX-2 may be the predominant source of prostaglandins in *H. pylori* gastritis, leading to an increased susceptibility to mucosal injury by COX-2 specific inhibitors. To date there are conflicting data showing that COX-2 specific inhibitors increase or have no effect on the risk of mucosal injury in the presence of *H. pylori* gastritis. How COX-2 specific inhibitors differ from nonselective NSAIDs in terms of their effects on *H. pylori*-infected gastric mucosa will be an interesting area of research.

### Expression and Cellular Localization of COX-2 in *H. pylori* Infection

Many studies have reported an increased expression of COX-2 in the presence of *H. pylori* infection. *H. pylori* has been shown to upregulate the expression of COX-2 messenger RNA (mRNA) and stimulates prostaglandin synthesis in gastric cancer cell lines [11]. However, there are conflicting data on the cellular localization of COX-2 expression in *H. pylori* gastritis. Fu *et al.* reported that *H. pylori* induces COX-2 expression in the mononuclear

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inflammatory cells and myofibroblasts in the lamina propria [10]. However, other studies found that COX-2 was expressed mainly in the gastric epithelium [1, 6, 7]. Sawaoka *et al.* reported that COX-2 was expressed both in the gastric epithelium and subepithelial inflammatory cells in *H. pylori* gastritis [9]. These inconsistent immunohistochemical findings probably are related to different laboratory conditions and cross-reactivity of COX-2 antibodies with other mucosal antigens. Using *in situ* hybridization, it has been shown that *H. pylori* up-regulates the expression of COX-2 mRNA mainly in the gastric epithelial cells [1].

#### Role of COX-1 and COX-2 in *H. pylori*-Induced Prostaglandin Synthesis

In the normal human stomach, COX-2 is absent or minimally expressed whereas COX-1 is the source of prostaglandins that maintains the integrity of the mucosal barrier. This notion is consistent with the observation that in the absence of *H. pylori* infection, inhibition of COX-2 did not suppress gastric prostaglandin synthesis and inflicted minimal mucosal injury [3]. However, in cultured human gastric fibroblasts [12, 13], *H. pylori* induced the expression of COX-2 mRNA and increased prostaglandin synthesis. Indomethacin and a COX-2 inhibitor (NS-398) suppressed *H. pylori*-induced prostaglandin synthesis to the same extent. These findings suggested that COX-2 may substantially contribute to prostaglandin synthesis in *H. pylori* gastritis; and that selective inhibition of COX-2 may lose its gastric sparing effect in the presence of *H. pylori* infection.

However, there were conflicting data on the relative contributions of COX-1 and COX-2 in prostaglandin synthesis associated with *H. pylori* gastritis. Jackson *et al.* [14] reported that COX-1 and COX-2 were constitutively expressed in parietal cells of uninfected human stomach. Immunostaining for both COX-1 and COX-2 was increased in *H. pylori* gastritis. Interestingly, the increased *ex vivo* prostaglandin synthesis was significantly suppressed by a COX-1 inhibitor rather than a COX-2 inhibitor. Scheiman *et al.* studied the effect of rofecoxib on gastric prostaglandin synthesis in subjects with or without *H. pylori* infection [15]. Twenty *H. pylori*-infected and 6 uninfected healthy volunteers were treated with rofecoxib for 2 weeks. Although prostaglandin levels were increased in *H. pylori* gastritis, rofecoxib did not suppress prostaglandin synthesis in infected subjects. These results suggested that despite an upregulation of COX-2, *H. pylori* gastritis, COX-1 remains the predominant source of prostaglandins.

#### EFFECTS OF NSAIDS AND COX-2 SPECIFIC INHIBITORS ON *H. pylori*-INFECTED GASTRIC MUCOSA

Gastric prostaglandins play a crucial role in mucosal defense by regulating mucosal blood flow, mucus and bicarbonate secretion, epithelial proliferation, epithelial restitution, and mucosal immunocyte function [16]. The fact that *H. pylori* infection stimulates gastric prostaglandin production has led to the speculation that *H. pylori* may alleviate mucosal injury induced by NSAIDs. However, administration of NSAIDs to *H. pylori*-infected subjects has been shown to profoundly suppress prostaglandin production to levels that were similar to those of uninfected subjects [17, 18]. These findings indicate that the modest increase in prostaglandin levels induced by *H. pylori* is unlikely to have any important protective effect against NSAID injury. It has been postulated that the mucosal toxicity of *H. pylori*, which is largely mediated by inflammatory cytokines including interleukin (IL)-1, IL-6, IL-8 and tumor necrosis factor, is counterbalanced by protective responses, such as increased release of mucosal prostaglandins and hepatocyte growth factor [12]. Factors disturbing this balance may enhance the gastric damaging effects of NSAIDs and mucosal toxicity of *H. pylori* [12]. To date there are only a few experimental and human studies investigating the effects of selective COX-2 inhibition on *H. pylori*-infected gastric mucosa.

#### Animal Studies (Table 1)

Several studies have investigated the effect of nonselective NSAIDs and a COX-2 specific inhibitor on *H. pylori* gastritis using the Mongolian gerbil model [13, 19, 20]. In one study [13], COX-1 was detected in both normal and *H. pylori*-infected mucosa whereas COX-2 was expressed only in the infected mucosa. *H. pylori* infection increased prostaglandin synthesis. Indomethacin markedly inhibited prostaglandin synthesis in both normal and infected mucosa. NS-398 also suppressed prostaglandin synthesis in infected mucosa but did not have any effect on uninfected mucosa. Hemorrhagic erosions and neutrophil infiltration were found in *H. pylori* gastritis. These mucosal lesions were aggravated by indomethacin and NS-398. Both drugs potentiated the release of neutrophil chemokine and interferon- $\gamma$  induced by *H. pylori*. In another study [19], indomethacin and NS-398 significantly suppressed gastric prostaglandin synthesis and there was a non-significant trend toward less severe suppression with NS-398 in *H. pylori*-infected gerbils. Indomethacin and NS-398 caused similar degree of gastric mucosal damage in infected animals despite

Table 1. Effects of NSAIDs and COX-2 Inhibitor on Animal Models of *H. pylori* Gastritis

Animal model	COX-1 & -2 expression	Baseline PGE2	PGE2 after NSAIDs	PGE2 after COX-2 inhibitor
Mongolian Gerbil [13]	COX-2 upregulated	Increased	Suppressed	Moderately suppressed*
Mongolian Gerbil [19]	---	Increased	Suppressed	Moderately suppressed†
Mouse [21]	COX-1 & -2 upregulated	Non-significant increase	Suppressed	Non-significant decrease

\*Mucosal damage was aggravated by both NSAIDs and NS-398 in *H. pylori*-infected mucosa.

†NSAIDs and NS-398 induced similar degree of mucosal damage in *H. pylori*-infected animals.

different degrees of prostaglandin suppression. In contrast, there was an inverse relationship between gastric prostaglandin level and mucosal damage in uninfected animals. These findings suggested that while COX-2 specific inhibitors caused minimal injury to uninfected gastric mucosa, these drugs did not reduce mucosal damage in *H. pylori* gastritis.

Unlike the previous two studies, Kim *et al.* [21] found that both COX-1 and COX-2 were upregulated in mouse stomachs infected with *H. pylori*. *H. pylori* infection increased apoptotic index, cell proliferation index, neutrophil activity and the degree of chronic inflammation. There was a non-significant increase in gastric prostaglandin levels. All these changes were reversed after the administration of indomethacin whereas NS-398 did not induce a significant reduction. The result suggested that both COX-1 and COX-2 are induced by *H. pylori* infection. Induction of COX-1 also contributes to the increase in prostaglandin synthesis, mucosal cell turnover and inflammatory activity in *H. pylori* gastritis.

Recently, Futagami *et al.* [20] investigated how inhibition of COX-2 would influence the severity of NSAID-induced gastric damage in *H. pylori*-infected Mongolian gerbils. *H. pylori* infection induced COX-2 expression. Prolonged treatment with indomethacin caused more severe gastric damage in *H. pylori*-infected animals than in uninfected animals. Interestingly, pretreatment with NS-398 aggravated the mucosal damage induced by short-term treatment with indomethacin in the presence of *H. pylori*. The authors postulated that induction of COX-2 by *H. pylori* might protect the gastric mucosa against NSAID injury. Disturbance of this equilibrium state by inhibiting COX-2 may enhance the gastric toxicity of NSAIDs in *H. pylori*-infected animals (Fig. 1).

#### Human Studies (Table 2)

Current evidence on the gastric safety of COX-2 inhibitors in *H. pylori*-infected patients is mostly derived

from *post hoc* analysis. Whether *H. pylori* infection increases the risk of ulcer disease in patients receiving COX-2 specific inhibitors has generated conflicting results in the literature.

The influence of *H. pylori* infection on the risk of gastroduodenal ulceration was first reported in a subgroup analysis of a double-blind, 12-week endoscopic study of celecoxib versus naproxen [22]. Among patients who received celecoxib, the incidence of ulcer was 12.9% in patients with *H. pylori* infection compared with 2.9% in uninfected patients ( $P=0.023$ ). Other risk factors included concurrent use of low-dose aspirin ( $P=0.001$ ) and a history of ulcer ( $P=0.010$ ). In contrast, *H. pylori* did not influence the risk of ulcer among patients who received naproxen. However, the same group of investigators reported contradictory results in a pooled analysis of four double-blind 12-week endoscopic studies of celecoxib that collectively enrolled 4000 arthritis patients [23]. Among patients who used nonselective NSAIDs, the incidence of ulcer was 28.4% in patients with *H. pylori* infection compared with 20% in uninfected patients (odds ratio 1.6 [1.1, 2.3]). Among patients who used celecoxib, the incidence of ulcer was 8.0% in patients with *H. pylori* infection compared with 5.1% in uninfected patients (odds ratio 1.6 [0.9, 2.8]). These results suggested that *H. pylori* is a risk factor for gastroduodenal ulceration in patients taking nonselective NSAIDs but not celecoxib.

In a multivariate analysis of risk factors for upper gastrointestinal clinical events [24] based on the data collected in the Vioxx Gastrointestinal Outcomes Research Study [5], major risk factors for the development of upper gastrointestinal clinical events included old age ( $\geq 75$ ) and prior complicated or uncomplicated gastrointestinal events. Patients with prior gastrointestinal events who received naproxen had a high rate of clinical events regardless of *H. pylori* status. Although *H. pylori* was not considered as a risk factor in this multivariate analysis, two interesting findings were reported. First, patients in the rofecoxib group had

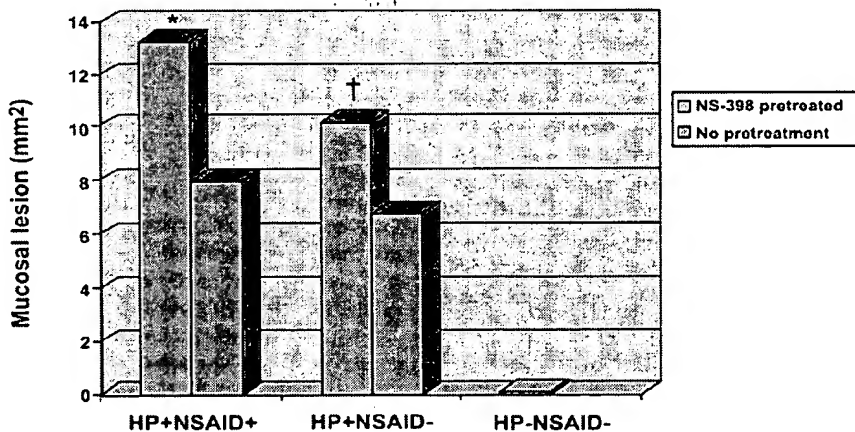


Fig. (1). Effects of Indomethacin on gastric damage in *H. pylori*-infected Mongolian gerbils with or without pretreatment with NS-398. HP and NSAID denote *H. pylori* and indomethacin, respectively. \* $P<0.05$  versus HP+NSAID+ group without pretreatment with NS-398. † $P<0.05$  versus HP+NSAID- group without pretreatment with NS-398. Data derived from [20].

Table 2. Clinical Effects of *H. pylori* Infection on Gastroduodenal Damage of COX-2 Specific Inhibitors

Design	Number of patients	Outcomes			
Subgroup analysis of a 12-week RCT of celecoxib 400 mg versus naproxen 1 g [22]	536	Endoscopic ulcer:			
			Celecoxib	Naproxen	
		HP positive	12.9%*	29%	
		HP negative	2.9%	30%	
		(P=0.023)*			
Pooled analysis of four 12-week RCTs of celecoxib 100-800 mg versus nonselective NSAIDs [23]	4000	Endoscopic ulcer (non-aspirin users):			
			Celecoxib	Nonselective NSAIDs	
		HP positive	8.0%	28.4%	
		HP negative	5.1%	20.0%	
		OR	1.6 (0.9 - 2.8)	1.6 (1.1 – 2.3)*	
Multivariate analysis of a RCT of rofecoxib 50 mg versus naproxen 1 g [24]	8076	Clinical upper GI events (per 100 patient-years):			
			Rofecoxib	Naproxen	
		Prior event, HP positive	12.18	14.00	RR 0.89 (0.38-2.07)†
		Prior event, HP negative	3.35	17.14	RR 0.20 (0.07-0.61)
		Complicated DU (per 100 patient-years):			
			Rofecoxib	Naproxen	
		HP positive	1.85	1.54	RR 1.20 (0.64-2.24)‡
		HP negative	0.34	1.41	RR 0.24 (0.09-0.64)

\**H. pylori* was a risk factor in patients taking nonselective NSAIDs but not in patients taking celecoxib

†The upper GI sparing effect of rofecoxib was offset by the presence of *H. pylori* infection in patients with prior upper GI events.

‡The superiority of rofecoxib over naproxen in reducing the risk of complicated duodenal ulcers disappeared in the presence of *H. pylori* infection.

fewer gastric ulcers than patients in the naproxen group regardless of the *H. pylori* status. In contrast, rofecoxib did not reduce the risk of duodenal ulcers compared with naproxen among patients found positive for *H. pylori*. Second, among those with prior gastrointestinal events, the rate of events in the rofecoxib group was 3.5-fold higher in *H. pylori*-positive patients than in *H. pylori*-negative patients. The results suggested that the upper GI sparing effect of rofecoxib was the offset by the presence of *H. pylori* infection in patients with prior upper GI events, and the superiority of rofecoxib over naproxen in reducing the risk of complicated duodenal ulcers disappeared in the presence of *H. pylori* infection.

#### EFFECTS OF *H. pylori* ON ULCER HEALING ASSOCIATED WITH NSAIDS AND COX-2 SPECIFIC INHIBITORS

Whether *H. pylori* infection affects ulcer healing in patients receiving nonselective NSAIDs has yielded conflicting data. In the rat model, one study found that *H. pylori* attenuated the delay in ulcer healing induced by aspirin. This observation was attributed to suppression of acid secretion and stimulation of prostaglandin production by *H. pylori* [25]. However, there are conflicting findings about the effects of *H. pylori* on aspirin-induced gastric injury. The same group of investigators showed that *H. pylori* induced persistent mucosal bleeding in the rat stomach by impairing gastric adaptation to aspirin [18]. Eradication of *H. pylori*

restores gastric adaptation to resist aspirin-induced injury. Hawkey *et al.* studied the effect of *H. pylori* eradication on gastroduodenal damage in chronic NSAID users with dyspepsia or ulcer [26]. In a subgroup of 41 patients with gastric ulcers, they found that eradication of *H. pylori* delayed ulcer healing (ulcer healing at 8 weeks: 72% in the eradicated group compared with 100% in the control group). In another randomized trial of *H. pylori*-positive patients with NSAID-associated ulcer bleeding, 195 patients (112 gastric ulcers and 83 duodenal ulcers) were randomly assigned to receive omeprazole or omeprazole plus eradication therapy for ulcer healing. Eradication of *H. pylori* did not have any significant adverse effect on the healing of gastric (*H. pylori*-positive versus *H. pylori*-eradicated: 94% versus 88%;  $p=0.29$ ) or duodenal (*H. pylori*-positive versus *H. pylori*-eradicated: 100% versus 98%;  $p=1.0$ ) ulcers [27] (Fig. 2). To date there is no definite evidence to show that eradication of *H. pylori* has any clinically important adverse effect on healing of NSAID-associated ulcers.

On the other hand, there are data suggesting that among patients receiving NSAIDs, gastric ulcers heal faster with *H. pylori* infection by acid suppression [28, 29]. *H. pylori* infection has been shown to augment the acid-suppressing effect of omeprazole [30, 31]. In a pooled analysis of three randomized trials of omeprazole for the prevention of mucosal injury in NSAID users [28], *H. pylori* appeared to enhance gastric ulcer healing by acid suppression but retard

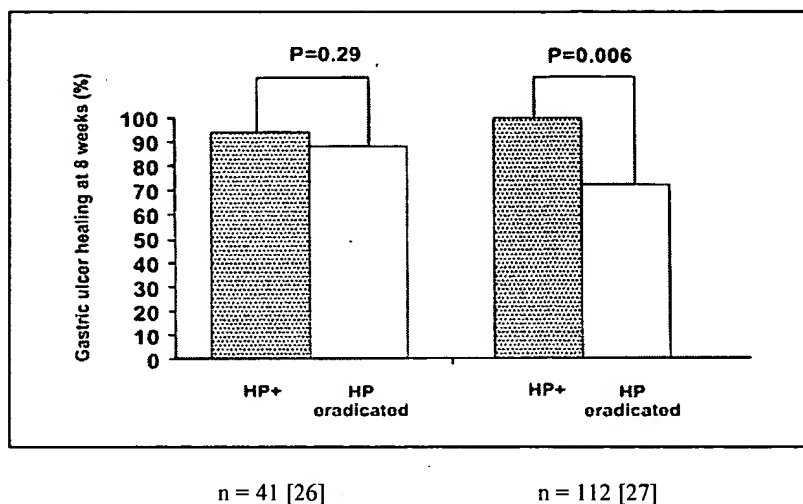


Fig. (2). Effects of *H. pylori* eradication on healing of gastric ulcers in patients receiving nonselective NSAIDs.

healing by misoprostol. However, the difference only reached statistical significance in patients receiving ranitidine (84% vs 51% at eight weeks) but not in patients receiving omeprazole. In another pooled analysis of two randomized trials of lansoprazole versus ranitidine in preventing NSAID-induced mucosal injury [29], gastric ulcer healing rates were significantly faster in *H. pylori*-positive patients than in uninfected patients though the difference was of doubtful clinical relevance (70% vs 61% at 8 weeks;  $P < 0.05$ ).

Although COX-2 specific inhibitors inflict minimal gastric injury to the normal stomach, there is evidence from animal experiments that COX-2 may play a physiological role in restoring mucosal integrity. In the rat stomach, COX-2 was upregulated in acetic acid-induced gastric ulcers [32, 33]. COX-2 activity was detected in endothelial cells, macrophages and fibroblasts at the ulcer base [32, 33]. Selective inhibition of COX-2 has been shown to retard gastric ulcer healing [33-36]. Administration of NS-398 significantly retarded healing of acetic acid-induced ulcers in rats and thermal-cauterized ulcers in mice [33-35]. One study showed that inhibition of COX-2-derived prostaglandins in ulcerated mucosa delayed ulcer healing [33]. Other investigators found that angiogenesis and maturation of granulation tissue in gastric ulcer was impaired by inhibition of COX-2 [36]. Jones *et al.* [37] demonstrated that both nonselective NSAIDs and COX-2 inhibitors acted on endothelial cells to inhibit angiogenesis. In contrast, COX-1 was absent [36] or not induced in ulcerated mucosa [33, 34].

Unlike rodent ulcers, To *et al.* [38] found that both COX-1 and COX-2 were upregulated in human gastric ulcers. At the ulcer margin, increased COX-1 expression was detected in lamina propria cells whereas COX-2 was strongly expressed in the hyperplastic foveolar epithelium. At the ulcer base, COX-1 and COX-2 were strongly expressed in myofibroblasts, macrophages and endothelial cells in the

granulation tissue. The findings were similar between *H. pylori* ulcers or NSAIDs ulcers. This raises the possibility that both isoforms of COX may contribute to ulcer healing in the human stomach regardless of the *H. pylori* status. However, other investigators found that although intense COX-2 immunoreactivity was detected in human gastric ulcers, there was no significant change in COX-1 expression in ulcerated mucosa [14, 39]. To date there is no clinical data as to whether COX-2 inhibitors would retard gastric ulcer healing.

## CONCLUSION

Current data on the interaction between *H. pylori* infection and selective COX-2 inhibition with respect to gastric damage are mostly derived from animal experiments or indirect clinical evidence based on *post hoc* analysis. Several interesting findings deserve further studying. In animal models of *H. pylori* gastritis, COX-2 inhibitors suppressed prostaglandin synthesis and aggravated mucosal damage. COX-1 appears to be the predominant source of prostaglandins in the human stomach albeit an upregulation of COX-2 in the presence of *H. pylori* infection [14, 15]. This may partly explain why *H. pylori* did not increase the risk of developing gastric ulcers among patients receiving rofecoxib [24]. However, the same study also indicated that rofecoxib did not reduce the risk of complicated duodenal ulcers in the presence of *H. pylori* infection. In addition, there was no advantage of rofecoxib over a nonselective NSAID for those with prior events and *H. pylori* infection in terms of the risk of clinical upper gastrointestinal events. Whether eradication of *H. pylori* will reduce the ulcer risk in these subgroups has not been investigated. In contrast, pooled analysis of data from randomized trials of celecoxib showed that *H. pylori* was a risk for ulcer disease in patients receiving nonselective NSAIDs but not in patients receiving celecoxib [23]. It is uncertain whether these contradictory

findings reflect differences in pharmacological properties, variations in study design or heterogeneity of *H. pylori*-infected patients. In animal models of gastric ulcer, COX-2 was upregulated in the granulation tissue and ulcer margin. Inhibition of COX-2 delayed healing of gastric ulcer in rodents. Limited data showed that COX-2 expression was increased in human gastric ulcer regardless of the *H. pylori* status. Whether inhibition of COX-2 will impair ulcer healing in the human stomach remains unknown. Future studies with pre-specified endpoints are needed to define the gastrointestinal risk of COX-2 inhibitors in different subgroups of *H. pylori*-infected patients.

## REFERENCES

References 40-42 are related articles recently published in *Current Pharmaceutical Design*.

- [1] Chan FK, To KF, Ng YP, Lee TL, Cheng AS, Leung WK et al. Expression and cellular localization of cyclooxygenase -1 and -2 in *Helicobacter pylori* gastritis. *Aliment Pharmacology Ther* 2001; 15: 187-194.
- [2] Chan FK. *Helicobacter pylori* and nonsteroidal anti-inflammatory drugs. *Gastroenterology Clin North Am* 2001; 30: 937-952.
- [3] Wight NJ, Gottesdiener K, Garlick NM, Atherton CT, Novak S, Gertz BJ, et al. Rofecoxib, a COX-2 inhibitors, does not inhibit human gastric mucosal prostaglandin production. *Gastroenterology* 2001; 120: 867-73.
- [4] Silverstein FE, Faich G, Goldstein JL, Simon LS, Pincus T, Whelton A, et al. Gastrointestinal toxicity with celecoxib vs nonsteroidal anti-inflammatory drugs for osteoarthritis and rheumatoid arthritis: the CLASS study: A randomized controlled trial. Celecoxib Long-term Arthritis Safety Study. *JAMA* 2000; 284: 1247-1255.
- [5] Bombardier C, Laine L, Reicin A, Shapiro D, Burgos-Vargas R, Davis B, et al. Comparison of upper gastrointestinal toxicity of rofecoxib and naproxen in patients with rheumatoid arthritis. *N Engl J Med* 2000; 343: 1520-1528.
- [6] McCarthy CJ, Crofford LJ, Greenon J, Scheiman JM. Cyclooxygenase-2 expression in gastric antral mucosa before and after eradication of *Helicobacter pylori* infection. *Am J Gastroenterology* 1999; 94: 1218-23.
- [7] McCartney SA, Mitchell JA, Fairclough PD, Farthing MJ, Warner TD. Selective COX-2 specific inhibitors and human inflammatory bowel disease. *Aliment Pharmacol Ther* 1999; 13: 115-117.
- [8] Reuter BK, Asfaha S, Buret A, Sharkey KA, Wallace JL. Exacerbation of inflammation-associated colonic injury in rat through inhibition of cyclooxygenase-2. *J Clin Invest* 1996; 98: 2076-2085.
- [9] Sawaoka H, Kawano S, Tsuji S, Tsuji M, Sun W, Gunawan ES, et al. *Helicobacter pylori* infection induces cyclooxygenase-2 expression in human gastric mucosa. *Prostaglandins Leukot Essent Fatty Acids* 1998; 59: 313-316.
- [10] Fu S, Ramanujam KS, Wong A, Fantry GT, Drachenberg CB, James SP, et al. Increased expression and cellular localization of inducible nitric oxide synthase and cyclooxygenase 2 in *Helicobacter pylori* gastritis. *Gastroenterology* 1999; 116: 1319-29.
- [11] Romano M, Ricci V, Memoli A, Tuccillo C, Di Popolo A, Sommi P, et al. *Helicobacter pylori* up-regulates cyclooxygenase-2 mRNA expression and prostaglandin E<sub>2</sub> synthesis in MKN 28 gastric mucosal cells in vitro. *J Biology Chem* 1998; 273: 28560-28563.
- [12] Takahashi M, Katayama Y, Takada H, Kuwayama H, Terano A. The effect of NSAIDs and a COX-2 specific inhibitor, on *Helicobacter pylori*-induced PGE<sub>2</sub> and HGF in human gastric fibroblasts. *Aliment Pharma Ther* 2000; 14; (Suppl 1), 44-49.
- [13] Takahashi S, Fujita T, Yamamoto A. Role of cyclooxygenase-2 in *Helicobacter pylori*-induced gastritis in Mongolian gerbils. *Am J Physiology Gastrointest Liver Physiology* 2000; 279: G791-G798.
- [14] Jackson LM, Wu KC, Mahida YR, Jenkins D, Hawkey CJ. Cyclooxygenase (COX) 1 and 2 in normal, inflamed, and ulcerated human gastric mucosa. *Gut* 2000; 47: 762-770.
- [15] Scheiman JM, Greenon JK, Lee J, Cryer B. Impact of COX-2 specific inhibition on human *Helicobacter pylori* (HP) Gastritis: Implications for ulcerogenesis and carcinogenesis. *Gastroenterology* 2002; 122: A87.
- [16] Wallace JL, Tigley AW. New insights into prostaglandins and mucosal defence. *Aliment Pharmacology Ther* 1995; 9: 227-235.
- [17] Laine L, Cominelli F, Sloane R, Casini-Raggi V, Marin-Sorensen M, Weinstein WM. Interaction of NSAIDs and *Helicobacter pylori* on gastroduodenal injury and prostaglandin production: a controlled double-blind trial. *Aliment Pharmacology Ther* 1995; 9: 127-35.
- [18] Konturek JW, Dembinski A, Konturek SJ, Stachura J, Domschke W. Infection of *Helicobacter pylori* in gastric adaptation to continued administration of aspirin in humans. *Gastroenterology* 1998; 114: 245-255.
- [19] Bhang CS, Lee HS, Kim SS, Song HJ, Sung YJ, Kim JI, et al. Effects of selective cyclooxygenase-2 inhibitor and non-selective NSAIDs on *Helicobacter pylori*-induced gastritis in Mongolian gerbils. *Helicobacter* 2002; 7: 14-21.
- [20] Futagami S, Hiratsuka T, Wada K, Tatsuguchi A, Tsukui T, Miyake K, et al. Inhibition of *Helicobacter pylori*-induced cyclooxygenase-2 aggravates NSAID-caused gastric damage in Mongolian gerbils. *Aliment Pharmacology Ther* 2002; 16: 847-855.
- [21] Kim TI, Lee YC, Lee KH, Han JH, Chon CY, Moon YM, et al. Effects of nonsteroidal anti-inflammatory drugs on *Helicobacter pylori*-infected gastric mucosae of mice: apoptosis, cell proliferation, and inflammatory activity. *Infect Immun* 2001; 69: 5056-63.
- [22] Goldstein JL, Correa P, Zhao WW, Burr AM, Hubbard RC, Verburg KM, et al. Reduced incidence of gastroduodenal ulcers with celecoxib, a novel cyclooxygenase-2 inhibitor, compared to naproxen in patients with arthritis. *Am J Gastroenterology* 2001; 96: 1019-1027.
- [23] Goldstein JL, Agrawal NM, Silverstein FE, Verburg KM, Burr AM, Hubbard RC, et al. Influence of *H. pylori* infection and/or low-dose aspirin on gastroduodenal ulceration in patients treated with placebo, celecoxib, or NSAIDs. *Gastroenterology* 1999; 116: A174.
- [24] Laine L, Bombardier C, Hawkey CJ, Davis B, Shapiro D, Brett C, et al. Stratifying the risk of NSAID-related upper gastrointestinal clinical events: results of a double-blind outcomes study in patients with rheumatoid arthritis. *Gastroenterology* 2002; 213: 1006-1012.
- [25] Konturek PC, Brzozowski T, Kwiecien S, Drozdowicz D, Harsch IA, Meixner H, et al. Effect of *Helicobacter pylori* on delay in ulcer healing induced by aspirin in rats. *Eur J Pharmacology* 2002; 451: 191-202.
- [26] Hawkey CJ, Tulassay Z, Szczepanski L, van Rensburg CJ, Filipowicz-Sosnowska A, Lanas A, et al. Randomised controlled trial of *Helicobacter pylori* eradication in patients on non-steroidal anti-inflammatory drugs: HELP NSAIDs study. *Lancet* 1998; 352: 1016-1021.
- [27] Chan FK, Sung JJ, Suen R, Lee YT, Wu JC, Leung WK, et al. Does eradication of *H. pylori* impair healing of nonsteroidal anti-inflammatory drug associated bleeding peptic ulcers? A prospective randomized study. *Aliment Pharmacology Ther* 1998; 12: 1201-1205.
- [28] Hawkey CJ, Wilson I, Naesdal J, Langstrom G, Swannell AJ, Yeomans ND. Influence of sex and *Helicobacter pylori* on development and healing of gastroduodenal lesions in non-steroidal anti-inflammatory drug users. *Gut* 2002; 51: 344-50.
- [29] Campbell DR, Haber MM, Sheldon E, Collis C, Lukasik N, Huang B, et al. Effect of *H. pylori* status on gastric ulcer healing in patients continuing nonsteroidal anti-inflammatory therapy and receiving treatment with lansoprazole or ranitidine. *Am. J. Gastroenterol.* 2002; 97: 2208-14.
- [30] Labenz J, Tillenburg B, Peitz U, Idstrom JP, Verdu EF, Stolte M, et al. *Helicobacter pylori* augments the pH-increasing effect of omeprazole in patients with duodenal ulcer. *Gastroenterology* 1996; 110: 725-732.
- [31] Gillen D, Wirz AA, Neithercut WD, Ardill JES, McCall KEL. *Helicobacter pylori* infection potentiates the inhibition of gastric acid secretion by omeprazole. *Gut* 1999; 44: 468-475.
- [32] Takahashi S, Shigeta J, Inoue H, Tanabe T, Okabe S. Localization of cyclooxygenase-2 and regulation of its mRNA expression in gastric ulcers in rats. *Am J Physiology* 1998; 275: G1137-G1145.

- [33] Shigeta J, Takahashi S, Okabe, S. Role of cyclooxygenase-2 in the healing of gastric ulcers in rats. *J Pharmacology Exp Ther* 1998; 286: 1383-1390.
- [34] Mizuno H, Sakamoto C, Matsuda K, Wada K, Uchida T, Noguchi H, et al. Induction of cyclooxygenase 2 in gastric mucosal lesions and its inhibition by the specific antagonist delays healing in mice. *Gastroenterology* 1997; 112: 387-397.
- [35] Ukawa H, Yamakuni H, Kato S, Takeuchi K. Effects of cyclooxygenase-2 selective and nitric oxide-releasing nonsteroidal antiinflammatory drugs on mucosal ulcerogenic and healing responses of the stomach. *Dig Dis Sci* 1998; 43: 2003-2011.
- [36] Schmassmann A, Peskar BM, Stettler C, Netzer P, Stroff T, Flogerzi B, et al. Effects of inhibition of prostaglandin endoperoxide synthase-2 in chronic gastro-intestinal ulcer models in rats. *Br J Pharmacology* 1998; 123: 795-804.
- [37] Jones MK, Wang H, Peskar BM, Levin E, Itani RM, Sarfeh IJ, et al. Inhibition of angiogenesis by nonsteroidal anti-inflammatory drugs: insight into mechanisms and implications for cancer growth and ulcer healing. *Nat Med* 1999; 5: 1418-1423.
- [38] To KF, Chan FK, Cheng AS, Lee TL, Ng YP, Sung JJ. Up-regulation of cyclooxygenase-1 and -2 in human gastric ulcer. *Aliment Pharmacology Ther* 2001; 15: 25-34.
- [39] Tatsuguchi A, Sakamoto C, Wada K, Akamatsu T, Tsukui T, Miyake K, et al. Localisation of cyclooxygenase 1 and cyclooxygenase 2 in *Helicobacter pylori* related gastritis and gastric ulcer tissues in humans. *Gut* 2000; 46: 782-789.
- [40] Hawkey CJ, Skelly MM. Gastrointestinal safety of selective COX-2 inhibitors. *Curr Pharm Design* 2002; 8(12): 1077-89.
- [41] Legrain P, Strosberg D. Protein interaction domain mapping for the selection of validated targets and lead compounds in the anti-infectious area. *Curr Pharm Design* 2002; 8(13): 1189-98.
- [42] Sobal G, Sinzinger H. Prostaglandins and lipid modification. *Curr Pharm Design* 2001; 7(6): 461-74.